



Neonatal liver physiology

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ABSTRACT

In the neonate, the liver is relatively immature and undergoes several changes in its functional capacity during the early postnatal period. The essential liver functions can be classified into three categories: metabolism, detoxification, and bile synthesis. In general, the immature liver function has limited consequences on the healthy term neonate. However, preterm neonates are particularly susceptible to the effects of the immature liver function placing them at risk of hypoglycemia, hyperbilirubinemia, cholestasis, bleeding, and impaired drug metabolism. An appreciation of the dynamic changes in liver function during the neonatal period is essential for successful management of neonates who require medical and surgical interventions. This review will focus on the neonatal liver function as well as the changes that the liver undergoes as it matures.

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Introduction

The liver is the largest solid organ and is responsible for several critical functions including the metabolism of dietary compounds, regulation of blood glucose levels, production of clotting factors and serum proteins, bile synthesis, and biotransformation of xenobiotics and endogenous by-products of metabolism.¹ During gestation, the fetus is able to rely on the metabolic activity of the maternal liver while the fetal liver develops the capacity to perform these functions. In the neonate, the liver is relatively immature and undergoes several changes in its functional capacity during the early postnatal period. In general, the immature liver function has limited consequences on the healthy term neonate. However, preterm neonates are particularly susceptible to the effects of the immature liver function placing them at risk of hypoglycemia, hyperbilirubinemia, cholestasis, bleeding, and impaired drug metabolism. An appreciation of the dynamic changes in liver function during the neonatal period is essential for successful management of neonates who require medical and surgical interventions. This review will focus on the neonatal liver function as well as the changes that the liver undergoes as it matures.

Liver embryology

Liver organogenesis begins during the fourth week of embryogenesis with the development of an outpouching from the ventral

foregut endoderm referred to as the hepatic diverticulum or liver bud. The cranial portion of the hepatic diverticulum develops into the liver and intrahepatic biliary tree while the caudal portion develops into the gallbladder and extrahepatic biliary tree. The primitive endodermal cells of this bud, also referred to as hepatoblasts, are bipotential cells with the ability to differentiate into hepatocytes and biliary epithelial cells (e.g., cholangiocytes).² The cardiac mesoderm and septum transversum are in close contact with hepatoblasts^{3,4} and secrete growth factors necessary for induction of hepatoblasts and differentiation and maturation of hepatocytes.^{5,6} Rapidly growing hepatoblasts migrate into the mesenchyme of the septum transversum eventually forming long cords of hepatocytes called hepatic plates. Hepatic plates are separated by mesenchymal cells that form the specialized porous endothelium of the sinusoids. The intrahepatic bile ducts are formed by the periportal hepatoblasts, which through interactions with neighboring cells and matrix proteins form a continuous single-layered ring around the portal mesenchyme and give rise to the bile ducts.^{7–9}

Fetal liver circulation

The fetal liver receives highly oxygenated blood via the umbilical vein which travels along the falciform ligament and connects to the left branch of the portal vein via the portal sinus.^{10,11} The umbilical vein supplies oxygen and nutrients to the fetal liver, supplying approximately 75% of the total blood to the liver.¹² The umbilical vein continues past the portal vein branches as the ductus venosus and delivers highly oxygenated blood to the inferior vena cava. The oxygenated blood is eventually directed across the patent foramen ovale to the left side of the heart.^{10,13}

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Thus, the ductus venosus acts as a shunt supplying oxygenated blood to the growing fetus. The degree of shunting through the ductus venosus is highest during early pregnancy when approximately 30% of the blood supplied by the umbilical vein is shunted through the ductus venosus. By 30 weeks of gestation, this value is reduced to 20%, resulting in approximately 80% of umbilical blood perfusing the liver. The ductus venosus is under adrenergic tonic control and vasodilates in the presence of nitric oxide and prostaglandins. During fetal hypoxemia, extensive dilation of the ductus venosus results in increased shunting to supply oxygenated blood to the heart and brain.¹⁴ The degree of shunting through the ductus venosus is also influenced by the hematocrit levels, umbilical venous pressure, and neural and endocrine regulation resulting in a dynamic variation in the degree of shunting. Closure of the ductus venosus occurs within the first 2 weeks of life in the majority of neonates. However, there is a significant delay in ductus venosus closure in preterm neonates and neonates that received antenatal corticosteroids.^{15,16}

Functional organization of the liver

The functional unit of the liver consists of the hepatic acinus which is developed by the third month of gestation.¹⁷ Each acinus includes a central hepatic vein connected to four to six portal triads via hepatic plates. Blood from the portal vein and hepatic artery enters the sinusoids which are located between hepatic plates before emptying into the central hepatic veins. Hepatocytes located within these functional units perform specific functions depending on their location within the acinus. Hepatocytes with similar functions are divided into three zones within the acinus. Hepatocytes closest to the portal triad are located in zone 1 and are exposed to blood with highest oxygen and nutrient concentration. As a result, these cells perform the majority of the liver's metabolic functions including glycogen synthesis, gluconeogenesis, protein synthesis, and lipid metabolism.^{18–22} The hepatocytes closest to the central veins are located in zone 3 and are specialized for biotransformation reactions, glycolysis, and urea synthesis. Hepatocytes in zone 2 are located between zones 1 and 3 and their function is dependent on their relative proximity to zone 1 or 3.

Plasma protein synthesis

Plasma protein synthesis and homeostasis is a vital function of the liver. The substrates necessary for plasma protein synthesis are provided by amino acids transported via the portal vein across the basolateral membrane of sinusoidal hepatocytes. The main serum protein produced by the fetal liver is alpha-fetoprotein which reaches its maximal concentration by the end of the first trimester. Albumin synthesis begins at approximately the 16th week of gestation and reaches adult levels by the end of gestation. All of the coagulation proteins are synthesized in the liver with the exception of factor VIII. The serum concentrations of coagulation proteins are low in newborn infants and reach adult levels within the first few days following birth.²³

Glucose and fatty acid metabolism

During gestation, the fetus receives a continuous supply of nutrients from the maternal circulation. The fetal glucose concentration is slightly lower, but in equilibrium with maternal glucose concentration and is the principle source of energy for the fetus under physiologic conditions. The fetal liver contains the necessary enzymes for gluconeogenesis and glycogen synthesis by the eighth

week of gestation. Gluconeogenesis does not occur *in utero* under physiologic conditions; however, glycogen synthesis begins early in pregnancy and glycogen deposition increases from 3.4 mg/g of liver tissue at 8 weeks gestational age to 50 mg/g of liver tissue by term.²⁴

Newborn infants have labile blood glucose levels as they transition from receiving nutrients from the maternal circulation to metabolizing hepatic stores and enteral feeds. At birth, the glucose concentration in umbilical venous blood is 80–90% of the maternal venous glucose concentration.²⁵ Blood glucose concentration in the neonate falls rapidly until 1 h of age after which glucose concentrations rise and stabilize by 3 h of age regardless of any enteral intake by the neonate.^{26,27} During this same period, plasma insulin levels fall and glucagon levels markedly increase.^{28,29} This resultant decrease in the insulin/glucagon ratio promotes mobilization of the limited hepatic glycogen stores which get reduced to 10% of their initial levels within 12 h of birth.³⁰ Following the depletion of hepatic glycogen stores, maintenance of normoglycemia in the neonate is dependent on metabolism of milk lactose provided enterally or through gluconeogenesis.

Extremely low blood glucose concentrations are uncommon in full-term breast-fed infants with modern feeding practices. The rate of glucose production in the neonate during the first few days following birth is estimated to be between 4 and 6 mg/kg/min.^{31,32} Glycogenolysis accounts for approximately one-third of this glucose production and gluconeogenesis supplies a significant proportion of the remainder. Fetal gluconeogenesis is limited secondary to low activity of phosphoenolpyruvate carboxykinase; however, following birth, the activity of this rate limiting enzyme markedly increases and results in the activation of gluconeogenesis as early as 2 h after birth.²⁴ In term neonates, hepatic glucose-6-phosphatase activity, the final enzymatic step in glycogenolysis and gluconeogenesis, is significantly reduced in the fetus, increasing to 10% of adult levels by term and reaching adult levels by 3 days of age.³³

Fatty acid metabolism provides another significant energy source for the neonate during the early postnatal period. Maternal plasma triglycerides undergo lipolysis in the placenta to allow for transport of fatty acids to the fetus. However, fetal fatty acid oxidation is low allowing for shuttling of fatty acids to adipose tissue for triglyceride synthesis.³⁴ Shortly after birth, the rate of lipolysis is increased and appears to be modulated by catecholamine release. During suckling, lipolysis is triggered due to enhanced sensitivity to lipolytic hormones (e.g., thyrotropin)³⁵ and decreased plasma insulin/glucagon ratio.^{36,37} Elevated lipolysis provides non-esterified fatty acids for oxidation and ketone body synthesis. Within the first day of life, hepatic ketogenesis is markedly elevated in term infants³⁸ and remains elevated during the first 3 days of life.³⁹ These findings suggest that ketone bodies may provide up to 25% of the neonate's basal energy requirements during the first few days of life.²⁴ Additionally, elevated hepatic ketogenesis has been shown to continue through the suckling period possibly due to a direct ketogenic effect of breast milk.³⁹

Preterm neonates have lower hepatic glycogen reserves⁴⁰ as well as decreased activity of gluconeogenic enzymes^{34,41} which result in lower postnatal glucose concentrations. During the first few hours following birth, there is a significantly greater decrease in glucose concentrations in preterm neonates compared to term neonates. This is due to lower hepatic glucose-6-phosphatase activity in preterm neonates. As a result, preterm neonates have a very limited response to exogenous glucagon administration which has been shown to stimulate hepatic glucose-6-phosphatase expression in animal studies.^{42,43} This illustrates the limited gluconeogenic capacity in premature neonates secondary to immaturity of responsible enzymatic pathways. In addition, serum ketone body levels are significantly lower in preterm

neonates indicating an inability to produce an appropriate ketogenic response to falling blood glucose levels in the early postnatal period. This immature hepatic ketogenic response to hypoglycemia in preterm infants may continue through the first 8 weeks of postnatal life.^{25,44} Therefore, maintenance of glucose levels in preterm neonates is complicated by reduced glycogen stores, immaturity of the gluconeogenic pathway, and an inadequate ketogenic response to hypoglycemia.

Bilirubin metabolism

Bilirubin is a by-product of hemoglobin catabolism from senescent red blood cells in the neonate. The heme moiety is metabolized to bilirubin within the reticuloendothelial system and transported to the liver while bound to albumin. Unconjugated bilirubin is highly lipophilic and water insoluble thus limiting its ability to be readily eliminated from the body. Conjugation of bilirubin with glucuronide is catalyzed by the hepatic enzyme uridine diphosphate glucuronyl transferase (UDPGT). Expression of hepatic UDPGT is approximately 1% of adult values between 30 and 40 weeks of gestation and increases significantly to adult levels during the first few weeks of life.⁴⁵ Conjugation of bilirubin increases its aqueous solubility and increases its transport into the canaliculus via the multispecific organic anion transport system. Conjugation of bilirubin also attenuates its toxic detergent effect on bile ducts.²³ Ultimately, conjugated bilirubin is excreted into bile and transported via the biliary tract to the small intestines where it is either excreted in stool or deconjugated by bacterial or enteric β -glucuronidase and reabsorbed into blood as part of the enterohepatic circulation.^{46,47}

A rise in unconjugated bilirubin during the first 2 weeks of life is very common and is usually a self-limiting process in the neonate. This physiologic jaundice results from increased bilirubin production, deficient conjugation, and/or increased reabsorption of unconjugated bilirubin via the enterohepatic circulation. During the first few days after birth, the neonate is not receiving full feeds and has not developed a normal stooling pattern. In addition, the hepatic uptake, conjugation, and biliary excretion of bilirubin are not functioning at their eventual full capacity.⁴⁸

Several factors are responsible for the increased severity and longer duration of hyperbilirubinemia observed in preterm neonates compared to term neonates.⁴⁸ Preterm neonates experience higher rates of bilirubin production due to the higher proportion of senescent red blood cells. The preterm neonatal liver also has a reduced ability to uptake bilirubin due to deficient organic anion transport proteins. Additionally, preterm neonates have reduced hepatic UDPGT activity resulting in decreased conjugation of bilirubin. Furthermore, feeding difficulties and increased conversion of conjugated bilirubin to its unconjugated form by enteric β -glucuronidase increases enterohepatic absorption of bilirubin. These factors result in an early onset, longer duration, and increased severity of hyperbilirubinemia in preterm infants.

Bile synthesis

Primary bile acids cholic acid and chenodeoxycholic acid are synthesized from cholesterol in the liver and are first detected in the fetus by the 14th week of gestation. Examination of bile present in meconium has revealed several distinguishing features between bile acids produced by fetal and adult livers. Fetal bile acids demonstrate an increased ratio of chenodeoxycholic acid to cholic acid which is in contrast to adults in whom there is a predominance of cholic acid. Another difference is the predominance of taurine conjugation in the fetus in contrast to glycine-conjugated bile acids

in adults. Finally, fetal bile acids have additional hydroxylations at carbons 1, 2, and 4 or 6 of the sterol nucleus.^{49–51} These findings demonstrate unique differences in the bile acid synthetic pathway between fetal and adult livers. The physiologic significance of these differences is not known but has been hypothesized to serve a protective role against hepatotoxic products produced by the fetal liver, such as lithocholate. In both preterm and term neonates, the bile acid pool is reduced; however, term neonates experience a significant expansion of the bile acid pool size at the end of gestation.^{52,53} By 7 weeks of age, the bile acid pool size in infants is comparable to adults when corrected for body surface area.⁵⁴

The molecular mechanisms responsible for generating bile flow and bile acid uptake are not fully developed in the neonate. Animal studies have been used to investigate the development of these processes in fetal and neonatal livers. Through these studies the developmental course of the enterohepatic circulation during fetal and early postnatal life has been demonstrated and provides some insight into the predisposition of neonates to cholestatic liver disease. These studies have demonstrated decreased bile flow and bile acid secretion in the developing liver.⁵⁵ During late gestation, there is increased activity of enzymes involved in bile acid synthesis resulting in an increase in bile acid pool size before birth.⁵⁶ At birth, bile acid transport across the basolateral and canalicular membranes of hepatocytes increases, resulting in a shift of bile acid pool from the liver to intestine.⁵² Additionally, the sodium–bile acid co-transport activity of the terminal ileum is fully developed by the time of weaning.⁵⁷ The final step in maturation of the enterohepatic circulation occurs when the hepatocyte sodium–bile acid co-transporter reaches its full potential, resulting in a decrease in serum bile acid concentration.⁵⁶ These findings demonstrate the development and maturation of the enterohepatic circulation in animal models; however, these findings have not been confirmed in human neonates. Shortly after birth, the serum bile acid levels in the neonate follow a characteristic developmental pattern. During the first week of life, the serum primary bile acids reach a significantly higher concentration than healthy older children and adults.⁵⁸ Additionally, serum bile acid levels do not decrease to levels comparable to those in adults until 6 months of age.⁵⁸ The high serum bile acid levels observed in infants during this time are the result of impaired hepatic uptake resulting in a physiologic cholestasis of infancy.^{59,60} The mechanisms for impaired hepatic uptake have been investigated in animal studies and revealed attenuated uptake of bile acids across the basolateral membrane of fetal and neonatal hepatocytes. This impaired hepatic bile acid uptake is correlated with decreased expression of the sodium–bile acid co-transporter mRNA, decreased sodium–bile acid co-transporter protein levels, and attenuated bile acid transport.⁶¹ Maturation of bile acid metabolism and the enterohepatic circulation occurs by the end of the first year of life.^{60,62}

Maturation of the neonatal gallbladder is necessary for efficient bile flow into the duodenum. Term neonates have more effective gallbladder function compared to preterm neonates demonstrated by larger fasting gallbladder volume and more effective gallbladder contractions.⁶³ The duodenal bile acid concentration in neonates following enteral administration of milk or magnesium sulfate is reduced compared to older children.⁶⁴ Additionally, in term neonates, bile secretion appears to be unresponsive to postprandial stimulation.⁶⁰ These findings indicate that the neonatal gallbladder function is not fully developed at birth.

Biotransformation

The liver plays a critical role in modulating the absorption, excretion, and metabolism of xenobiotics and endogenous by-products of metabolism. During the neonatal period, the main

limiting factor affecting hepatic clearance of these products is either an immature drug metabolism pathway or hepatic transport mechanisms.⁶⁵ The role of membrane transporters in hepatic clearance of xenobiotics is poorly understood; however, limited investigation has demonstrated that the hepatic excretory function is immature in neonates.⁶⁶

Hepatic biotransformation is generally divided into phase I and phase II reactions. Phase I or activation reactions involve oxidation, reduction, and hydrolytic reactions. The principal enzymes involved in these activation reactions belong to the cytochrome P450 family. Phase II or detoxification reactions involve the conjugation of a water-soluble endogenous molecule (e.g., glucuronic acid, sulfate, glycine, glutathione, and acetate) in order to catalyze the metabolism of endogenous compounds or xenobiotics. Many of the enzymes involved in phase I and II reactions are differentially expressed during the perinatal period and subject to developmental alterations.

The neonatal liver undergoes a rapid maturation process during the first year of life which affects its many biotransformation functions. The cytochrome P450 enzymes are members of a superfamily that catalyze the oxidative metabolism of lipophilic substrates. The cytochrome P450 families 1, 2, and 3 play an important role in drug metabolism in the liver and are expressed early in fetal life.⁶⁵ The protein concentrations of cytochrome P450 enzymes remain relatively stable throughout gestation and are at 30% of adult concentrations at birth.^{67,68} These findings are not surprising given their role in modulating levels of endogenous substrates involved in fetal homeostasis, growth, and differentiation.⁶⁹ Individual cytochrome P450 enzymes demonstrate significant differences in expression and activity in the fetus and neonate compared to adults, resulting in vastly different metabolic profiles⁶⁵ between neonates and adults. During the postnatal period, total hepatic cytochrome P450 expression increases, reaching adult levels by 1 year of age.⁶⁷ These changes in cytochrome P450 expression during the first year of life result in decreased half-lives and increased clearance of certain drugs.^{70,71}

Phase II reactions increase the water solubility of xenobiotics or endogenous compounds for renal or biliary excretion through conjugation reactions. Important conjugation reactions in the infant include glucuronidation, sulfation, acetylation, glutathione conjugation, methylation, and amino acid conjugation.⁶⁵ In general, the developmental pattern and degree of fetal and neonatal expression of phase II enzymes is enzyme specific.⁷² Neonates have limited enzymatic glucuronidation capacity as demonstrated by their limited ability to conjugate bilirubin during early postnatal life resulting in unconjugated hyperbilirubinemia. These observations are consistent with the finding of low expression levels of hepatic UDPGT enzymes during fetal and early postnatal development. UDPGT expression increases after birth, reaching about 25% of adult levels by 3 months of age.⁷³ For example, both term and preterm infants demonstrate reduced and variable morphine clearances secondary to immature glucuronidation pathways.⁷⁴ Another important group of phase II enzymes are the sulfotransferases that catalyze sulfate conjugation of xenobiotics and endogenous products. Investigation of hepatic sulfotransferase activity in the fetus and neonates revealed high activity of this enzymatic pathway suggesting that it may have an essential role in homeostasis and detoxification in the fetus and neonate.⁶⁵

The hepatic drug metabolism pathways in the neonate are in general immature, resulting in prolonged drug elimination and increased plasma half-lives. However, the extent of compromised drug elimination in the neonate depends on the maturity of the metabolic pathway for a particular substrate and alternative routes of elimination. It should be noted that significant inter-individual variability in maturation of these metabolic pathways does exist.

Summary

The liver is undergoing dynamic changes during the neonatal period as it continues to mature. Immediately after birth, it has a vital role in glucose and fatty acid metabolism to ensure a continuous supply of energy as the neonate adapts to enteral feeding. It is responsible for the synthesis of plasma proteins and coagulation factors. In addition, it is responsible for bilirubin metabolism and bile synthesis for the elimination of metabolic wastes and absorption of essential nutrients as well as metabolism and clearance of xenobiotics. The maturation of the liver in term neonates is an expected natural process. However, due to the delay in liver maturation of preterm neonates, they have a significantly lower capacity to respond to the various environmental and physiologic demands. An appreciation of neonatal liver function is essential to the management of neonates undergoing medical and surgical interventions.

References

- Diehl-Jones WL, Askin DF. The neonatal liver. Part 1: embryology, anatomy and physiology. *Neonatal Netw.* 2002;21(2):5–12.
- Shiojiri N. Development and differentiation of bile ducts in the mammalian liver. *Microsc Res Tech.* 1997;39(4):328–335.
- Zaret KS. Hepatocyte differentiation: from the endoderm and beyond. *Curr Opin Genet Dev.* 2001;11(5):568–574.
- Duncan SA. Mechanisms controlling early development of the liver. *Mech Dev.* 2003;120(1):19–33.
- Rossi JM, Dunn NR, Hogan BL, Zaret KS. Distinct mesodermal signals, including BMPs from the septum transversum mesenchyme, are required in combination for hepatogenesis from the endoderm. *Genes Dev.* 2001;15:1998–2009.
- Houssaint E. Differentiation of the mouse hepatic primordium. I. An analysis of tissue interactions in hepatocyte differentiation. *Cell Differ.* 1980;9:269.
- Amenta PS, Harrison D. Expression and potential role of the extracellular matrix in hepatic ontogenesis: a review. *Microsc Res Tech.* 1997;39:372.
- Shah KD, Gerber MA. Development of intrahepatic bile ducts in humans. Possible role of laminin. *Arch Pathol Lab Med.* 1990;114:597.
- Terada T, Nakanuma Y. Expression of tenascin, type IV collagen and lamin during human intrahepatic bile duct development and intrahepatic cholangiocarcinoma. *Histopathology.* 1994;25:143.
- Rudolph AM. Hepatic and ductus venosus blood flows during fetal life. *Hepatology.* 1983;3:254.
- Nagano K, Hoshino H, Nishimura D, Katada N, Sano H, Kato K. Patent ductus venosus. *J Gastroenterol Hepatol.* 1999;14(3):285–288.
- Haugen G, Hanson M, Kiserud T, Crozier S, Inskip H, Godfrey KM. Fetal liver-sparing cardiovascular adaptations linked to mother's slimmness and diet. *Circ Res.* 2005;96(1):12–14.
- Edelstone DL. Regulation of blood flow through the ductus venosus. *J Dev Physiol.* 1980;2(4):219.
- Behrman RE, Lees MH, Peterson EN, De Lannoy CW, Seeds AE. Distribution of the circulation in the normal and asphyxiated fetal primate. *Am J Obstet Gynecol.* 1970;108(6):956–969.
- Kondo M, Itoh S, Kunikata T, et al. Time of closure of ductus venosus in term and preterm neonates. *Arch Dis Child Fetal Neonatal Ed.* 2001;85(1):F57–F59.
- Loberant N, Herskovits M, Barak M, et al. Closure of the ductus venosus in premature infants: findings on real-time gray-scale, color-flow doppler, and duplex doppler sonography. *Am J Roentgenol.* 1999;172(1):227–229.
- Pineiro-Carrero V, Pineiro E. Liver. *Pediatrics.* 2004;113(4):1097–1106.
- Tanaka T, Watanabe J, Asaka Y, Ogawa R, Kanamura S. Quantitative analysis of endoplasmic reticulum and cytochrome P-450 in hepatocytes from rats injected with methylcholanthrene. *Eur J Cell Biol.* 1997;74(1):20–30.
- Aggarwal SR, Lindros KO, Palmer TN. Glucagon stimulates phosphorylation of different peptides in isolated periportal and perivenous hepatocytes. *FEBS Lett.* 1995;377(3):439–443.
- McCashland RM, Tuma DJ, Sorrell MR, Casey CA. Zonal differences in ethanol-induced impairments in hepatic receptor binding. *Alcohol.* 1993;10(6):549–554.
- Lawrence GM, Trayer IP, Walker DG. Histochemical and immunohistochemical localization of hexokinase isoenzymes in normal rat liver. *Histochem J.* 1984;16(10):1099–1111.
- Hildebrand R. Quantitative and qualitative histochemical investigation on NADP⁺-dependent dehydrogenases in the limiting plate and the residual parenchyma surrounding terminal hepatic venules. *Histochemistry.* 1984;80(1):91–95.
- Beath SV. Hepatic function and physiology in the newborn. *Semin Neonatol.* 2003;8:337–346.
- Platt MW, Deshpande S. Metabolic adaptation at birth. *Semin Fetal Neonatal Med.* 2005;10:341–350.

25. Kalhan SC, Bier DM, Savin SM, Adam PA. Estimation of glucose turnover and ¹³C recycling in the human newborn by simultaneous [^{1-¹³C}] glucose and [6-6-²H₂] glucose tracers. *J Clin Endocrinol Metab.* 1980;50(3):456–460.
26. Heck LJ, Erenberg A. Serum glucose levels in term neonates during the first 48 hours of life. *J Pediatr.* 1987;110(1):119–122.
27. Srinivasan G, Pildes RS, Cattamanchi G, Voora S, Lillien LD. Plasma glucose values in normal neonates: a new look. *J Pediatr.* 1986;109(1):114–117.
28. Bloom SR, Johnston DI. Failure of glucagon release in infants of diabetic mothers. *Br Med J.* 1972;4(5838):453–454.
29. Sperling MA, DeLamater PV, Phelps D, Fiser RH, Oh W, Fisher DA. Spontaneous and amino acid stimulated glucagon secretion in the immediate post-natal period: relation to glucose and insulin. *J Clin Invest.* 1974;53(4):1159–1166.
30. Shelley HJ. Glycogen reserves and their changes at birth and in anoxia. *Br Med Bull.* 1961;17:137–143.
31. Kalhan SC, Savin SM, Adam PA. Measurement of glucose turnover in the human newborn with glucose-1-¹³C. *J Clin Endocrinol Metab.* 1976;43(3):704–707.
32. Bier DM, Leake RD, Haymond MW, et al. Measurement of 'true' glucose production rates in infancy and childhood with 6,6-dideuteroglucose. *Diabetes.* 1977;26(11):1016–1023.
33. Burchell A, Gibb L, Waddell ID, Giles M, Hume R. The ontogeny of the human hepatic glucose-6-phosphatase proteins. *Clin Chem.* 1990;36(9):1633–1637.
34. Herrera E, Amusquivar E. Lipid metabolism in the fetus and newborn. *Diabetes Metab Res Rev.* 2000;16(3):202–210.
35. Marcus C, Ehren H, Bolme P, Arner P. Regulation of lipolysis during the neonatal period: importance of thyrotropin. *J Clin Invest.* 1988;82(5):1793–1797.
36. Girard JR, Cuendet GS, Marliss EB, Kervran A, Rieutort M, Assan R. Fuels, hormones and liver metabolism at term and during the early postnatal period in the rat. *J Clin Invest.* 1973;52(12):3190–3200.
37. Issad T, Coupe C, Ferre P, Girard J. Insulin resistance during suckling period in rats. *Am J Physiol.* 1987;253(2, Pt 1):E142–E148.
38. Bougneres PF, Lemmel P, Ferre P, Bier DM. Ketone body transport in the human neonate and infant. *J Clin Invest.* 1986;77(1):42–48.
39. Hawdon J, Ward Platt M, Aynsley-Green A. Patterns of metabolic adaptation for preterm and term infants in the first neonatal week. *Arch Dis Child.* 1992; 67(4 Spec No):357–365.
40. Shelley HJ. Glycogen reserves and their changes at birth. *Br Med Bull.* 1975;3:137–143.
41. Hume R, Burchell A. Abnormal expression of glucose-6-phosphatase in preterm infants. *Arch Dis Child.* 1993;68(2):202–204.
42. Jackson L, Burchell A, McGeechan A, Hume R. An inadequate glycaemic response to glucagon is linked to insulin resistance in preterm infants? *Arch Dis Child.* 2003;88(1):F62–F66.
43. Greengard O, Dewey HK. Initiation by glucagon of the premature development of tyrosine aminotransferase, serine dehydratase, and glucose-6-phosphatase in fetal rat liver. *J Biol Chem.* 1967;242(12):2986–2991.
44. Deshpande S, Hawdon JM, Ward Platt MP, Aynsley-Green A. Metabolic adaptation to extrauterine life. In: Rodeck CH, Whittle MJ, editors. *Fetal Medicine: Basic Science and Clinical Practice.* London: Churchill Livingstone; 1999. p. 1059–1069.
45. Watchko JF, Lin Z. Exploring the genetic architecture of neonatal hyperbilirubinemia. *Semin Fetal Neonatal Med.* 2010;15(3):169–175.
46. Dennery PA, Seidman DS, Stevenson DK. Neonatal hyperbilirubinemia. *N Engl J Med.* 2001;344(8):581–590.
47. Watchko JF, Maisels MJ. Jaundice in low birth weight infants: pathobiology and outcome. *Arch Dis Child Fetal Neonatal Ed.* 2003;88(6):F455–F458.
48. Raju TNK. Developmental physiology of late and moderate prematurity. *Semin Fetal Neonatal Med.* 2012;17(3):126–131.
49. Lester, St Pyrek J, Little JM, Adcock EW. Diversity of bile acids in the fetus and newborn infant. *J Pediatr Gastroenterol Nutr.* 1983;2(2):355–364.
50. Setchell KD, Dumaswala R, Colombo C, Ronchi M. Hepatic bile acid metabolism during early development revealed from the analysis of human fetal gallbladder bile. *J Biol Chem.* 1988;263(32):16637–16644.
51. Gustafsson J. Bile acid biosynthesis during development: hydroxylation of C-27 sterols in human fetal liver. *J Lipid Res.* 1986;27(8):801–806.
52. Little JM, Richey JE, Van Thiel DH, Lester R. Taurocholate pool size and distribution in the fetal rat. *J Clin Invest.* 1979;63(5):1042–1049.
53. Subbiah MT, Hassan AS. Development of bile acid biogenesis and its significance in cholesterol homeostasis. *Adv Lipid Res.* 1982;19:137–161.
54. Heubi JE, Balistreri WF, Suchy FJ. Bile salt metabolism in the first year of life. *J Lab Clin Med.* 1982;100(1):127–136.
55. Shaffer EA, Zahazi I, Gall DG. Postnatal development of hepatic bile formation in the rabbit. *Dig Dis Sci.* 1985;30(6):558–563.
56. Henning SJ. Postnatal development: coordination of feeding, digestion, and metabolism. *Am J Physiol.* 1981;241(3):G199–G214.
57. Barnard JA, Ghishan FK, Wilson FA. Ontogenesis of taurocholate transport by rat ileal brush border membrane vesicles. *J Clin Invest.* 1985;75(3):869–873.
58. Suchy FJ, Balistreri WF, Heubi JE, Searcy JE, Levin RS. Physiologic cholestasis: elevation of the primary serum bile acid concentrations in normal infants. *Gastroenterology.* 1981;80(5 Pt 1):1037–1041.
59. Lester R. Physiologic cholestasis. *Gastroenterology.* 1980;78(4):864–865.
60. Balistreri WF, Heubi JE, Suchy FJ. Immaturity of the enterohepatic circulation in early life: factors predisposing to "physiologic" maldigestion and cholestasis. *J Pediatr Gastroenterol Nutr.* 1983;2(2):346–354.
61. Hardikar W, Ananthanarayanan M, Suchy FJ. Differential ontogenic regulation of basolateral and canalicular bile acid transport proteins in rat liver. *J Biol Chem.* 1995;270(35):20841–20846.
62. Heubi JE, Balistreri WF, Suchy FJ. Bile salt metabolism in the first year of life. *J Lab Clin Med.* 1982;100(1):127–136.
63. Ho ML, Chen JY, Ling UP, Su PH. Gallbladder volume and contractility in term and preterm neonates: normal values and clinical applications in ultrasonography. *Acta Paediatr.* 1998;87(7):799–804.
64. Ricour C, Rey J. Study of the hydrolysis and micellar solubilization of fats during intestinal perfusion. I. Results in the normal child. *Rev Eur Etud Clin Biol.* 1972;17(2):172–178.
65. Alcorn J, McNamara PJ. Ontogeny of hepatic and renal systemic clearance pathways in infants Part 1. *Clin Pharmacokinet.* 2002;41(12):959–998.
66. Balistreri WF. Immaturity of hepatic excretory function and the ontogeny of bile acid metabolism. *J Pediatr Gastroenterol Nutr.* 1983;2(suppl 1):S207–S214.
67. Treluyer JM, Cheron G, Sonnier M, Cresteil T. Cytochrome P-450 expression in sudden infant death syndrome. *Biochem Pharmacol.* 1996;52(3):497–504.
68. Treluyer JM, Gueret G, Cheron G, Sonnier M, Cresteil T. Developmental expression of CYP2C and CYP2C-dependent activities in the human liver: *in-vivo/in-vitro* correlation and inducibility. *Pharmacogenetics.* 1997;7(6):441–452.
69. Nebert DW. Proposed role of drug-metabolizing enzymes: regulation of steady state levels of the ligands that effect growth, homeostasis, differentiation, and neuroendocrine functions. *Mol Endocrinol.* 1991;5(9):1203–1214.
70. Gilman JT. Therapeutic drug monitoring in the neonate and paediatric age group: problems and clinical pharmacokinetic implications. *Clin Pharmacokinet.* 1990;19(1):1–10.
71. Sato Y. Pharmacokinetics of antibiotics in neonates. *Acta Paediatr Jpn.* 1997; 39(1):124–131.
72. Krauer B, Dayer P. Fetal drug metabolism and its possible clinical implications. *Clin Pharmacokinet.* 1991;21(1):70–80.
73. Coughtrie MW, Burchell B, Leakey JE, Hume R. The inadequacy of perinatal glucuronidation: immunoblot analysis of the developmental expression of individual UDP-glucuronosyltransferase isoenzymes in rat and human liver microsomes. *Mol Pharmacol.* 1988;34(6):729–735.
74. Mikkelsen S, Feilberg VL, Christensen CB, Lundstrom KE. Morphine pharmacokinetics in premature and mature newborn infants. *Acta Paediatr.* 1994;83(10): 1025–1028.